

12, 93349-38-3; 12-H₂C₂O₄, 93349-47-4; 13, 93349-39-4; 13-H₂C₂O₄, 93349-48-5; 14, 93349-40-7; 14-H₂C₂O₄, 93349-49-6; 15, 93349-41-8; 15-H₂C₂O₄, 93349-50-9; 16, 36125-06-1; 17, 83495-51-6; 18, 3897-89-0; 19, 619-73-8; 20, 22114-98-3; 21, 13605-19-1; 3-pentanone

oxime, 1188-11-0; cyclopentanone oxime, 1192-28-5; cyclohexanone oxime, 100-64-1; 4-heptanone oxime, 1188-63-2; 2,4-dimethyl-3-pentanone oxime, 1113-74-2; isopropylamine, 75-31-0; *tert*-butylamine, 75-64-9.

Syntheses and Complement Inhibitory Activities of 4-(2-Phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic Acids

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The syntheses of 4-(2-phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic acids are described. These compounds express potent *in vitro* inhibition of the human classical complement pathway, and qualitative SAR have been determined. Several of the *in vitro* active compounds also suppressed the complement dependent reverse passive Arthus reaction (RPAR) in guinea pigs.

The potential utility for drugs that inhibit complement dependent acute inflammatory events in diseases such as rheumatoid arthritis,² lupus erythematosus,³ and glomerulonephritis⁴ has been well documented. Although many chemicals, including several antiinflammatory agents, have been reported to be inhibitory of the complement cascade *in vitro*,⁵ none has been shown to display clinical efficacy via this mechanism. Thus, with clear relevance to disease, an investigation was initiated to discover a complement inhibitory drug that would have utility in the treatment of acute inflammatory disease. It is the purpose of this paper to report on results of this effort by describing the syntheses and potent classical complement pathway inhibitory properties of a series of 4-(2-phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic acids (5-18).

Chemistry. Scheme I depicts the several routes used to prepare the 4-(2-phenyl-1*H*-indol-3-yl)cyclohexane-carboxylic acids 5-18. Central in this scheme is the AcOH/Ac₂O/H₃PO₄ condensation⁶ of 2-phenyl-1*H*-indole (1) or 1-methyl-2-phenyl-1*H*-indole (2) with either 4-oxo-1,1-cyclohexanedicarboxylic acid (3) or 4-oxocyclohexanecarboxylic acid (4) that produced 5, 6, or 7.

The reaction of 5 with MeOH/MeSO₃H or MeI/K₂CO₃/DMF gave the 5 Me ester. Similarly, 6 diMe ester or 7 diMe ester were formed upon treatment of 6 or 7 with MeI/K₂CO₃/DMF. N-Alkylation of 5 Me ester with MeI/KOH/Me₂SO⁷ gave 8 Me ester, which was hydrolyzed with aqueous KOH to give 8. N-Alkylation products 9 and 10 were prepared by alkylation of 5 with excess EtI or PrBr and NaH in DMF followed by KOH hydrolysis of the intermediate esters. The EtI alkylation procedure with NaH followed by alkaline hydrolysis was also used to prepare 11 from 6 diMe ester.

Hydrogenation of diacids 6 or 7 over Pd/C gave directly 12 and 13, respectively. Hydrogenation products of monoacid 8 were prepared by a circuitous route which involved hydrogenation of 8 Me ester to give both *cis* (predominant) and *trans* (minor) products which were separated and hydrolyzed to the acids 14 and 15. Stereochemical assignments of *cis* for 14 and 14 Me ester, and

Table I. Effect of 4-(2-Phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic Acids on the Classical Complement Pathway in Vitro and the Reverse Passive Arthus Reaction (RPAR) in Guinea Pigs^a

compd	classical complement pathway inhibn, IC ₅₀ ^b	RPAR, inhibn (dose ^c)
5	135 ± 2	64 ± 7 ^d (80 iv)
6	485 ± 17	38 ± 8 ^e (100 ip)
7	149 ± 25	
8	34 ± 3	78 ± 6 ^e (300 ip)
9	65 ± 5	
10	93 ± 44	
11	130 ± 20	
12	425 ± 105	
13	231 ± 7	
14	>1000	66 ± 5 ^e (100 ip)
15	42 ± 21	55 ± 8 ^d (100 ip)
16	51 ± 15	76 ± 5 ^d (100 ip)
17	105 ± 4	
18	51 ± 11	44 ± 14 ^f (100 ip)
gold sodium thiomalate (GST)	1070 ± 90	
cobra venom factor (CVF)		ED ₅₀ = 7.9 units/kg ip

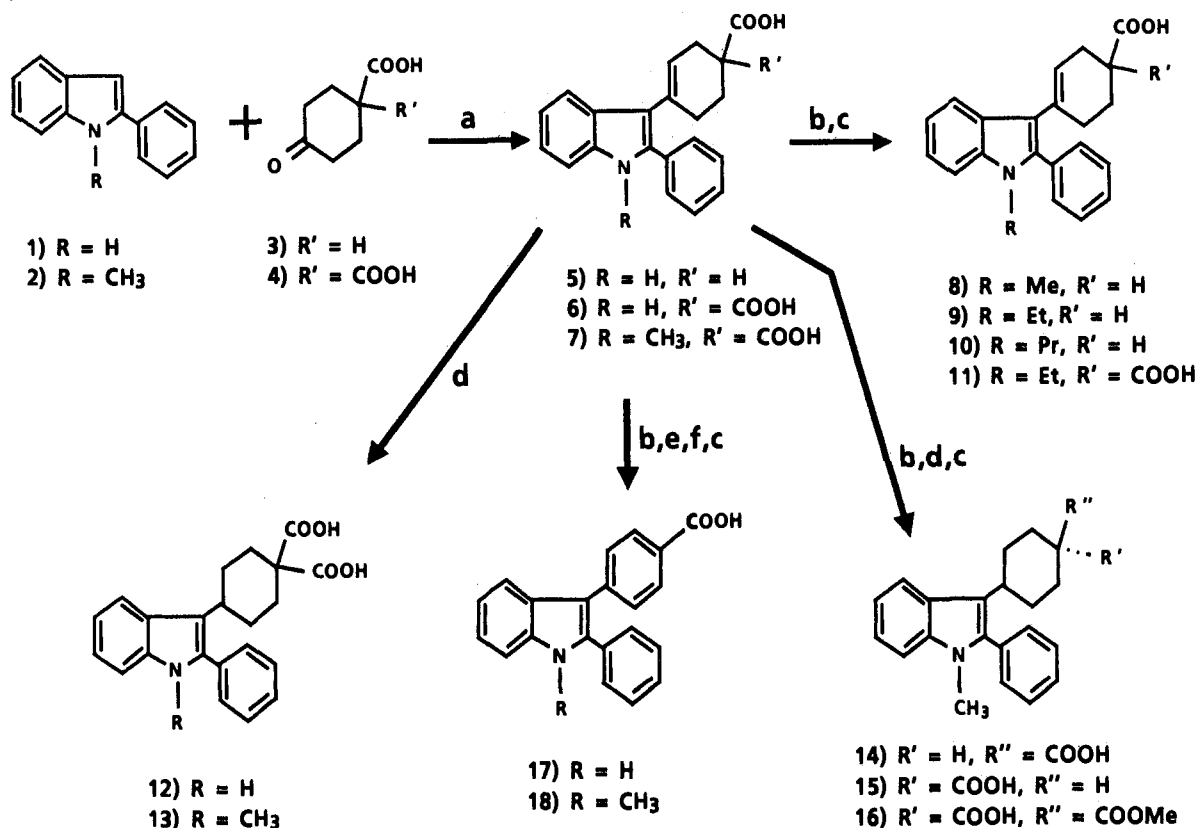
^a See Experimental Section for details of assay systems. ^b IC₅₀ in μM. ^c Dose in mg/kg. ^d *p* < 0.001. ^e *p* < 0.01. ^f *p* < 0.025.

trans for 15 and 15 Me ester were based on the reported higher ¹³C NMR cyclohexane methylene absorption frequencies for *cis* relative to *trans* of 1,4-dimethylcyclohexane⁸ and 4-(1,1-dimethylethyl)-1-(methylsulfinyl)-

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Scheme I^a

^a (a) H₃PO₄, AcOH, Ac₂O. (b) Base, RX. (c) KOH. (d) H₂, Pd/C. (e) K₂CO₃, MeI. (f) DDQ.

cyclohexane⁹. Thus, *cis*-14 and *cis*-14 Me ester have C_{2,6} absorptions at δ 27.3 and 27.8 and C_{3,5} absorptions at δ 28.6 and 29.1, respectively, while *trans*-15 and *trans*-15 Me ester have C_{2,6} absorptions at δ 31.6 and 32.3 and C_{3,5} absorptions at δ 29.1 and 29.4, respectively. Hydrogenation of 7 di Me ester followed by mild basic hydrolysis gave a single monoacid-monoester 16 which was assigned *trans* acid stereochemistry because equatorial cyclohexanecarboxylic acid esters are known to be more rapidly hydrolyzed than their axial isomers.¹⁰

Aromatization of 5 Me ester or 8 Me ester with DDQ followed by alkaline hydrolysis gave benzoic acids 17 and 18, respectively.

Biology. Compounds 5–18 were evaluated *in vitro* for their inhibition of human complement (C1–C9) using hemolytic molecular titrations¹¹ (Table I), and comparative IC₅₀s were calculated from the resulting dose responses. The IC₅₀ calculated for gold sodium thiomalate (GST), a known complement inhibitor,¹² was 1070 μ M and has been included in Table I for reference purposes. Thirteen of the 14 carboxylic acids tested by this method exhibited potent inhibition of the classical complement pathway (Table I) with a range in IC₅₀ from 34 (8) to 485 μ M (6). Correlation of the IC₅₀s with structure has led to the identification of several structural features that enhance potency. The most important feature is the *trans* stereochemistry of the carboxylic acid relative to the indole on

the cyclohexane ring which allows the indole and carboxylic acid moieties to be fully extended and maximally separated from each other. Compound 15 has this *trans* stereochemistry and has an IC₅₀ = 42 μ M, while 14, which has the *cis* stereochemistry, has an IC₅₀ > 1000 μ M. The *trans* carboxylic acid stereochemistry is also present in inhibitors 12, 13, and 16. Complement inhibitory, quasi-equatorial cyclohexanecarboxylic acids 5–11 and benzoic acids 17 and 18 have carboxylic acids that are positioned relative to the indole ring in an orientation similar to the *trans* cyclohexanes. The distance between the 3-indole carbon and the carboxylic acid for *trans* cyclohexane, cyclohexene, and benzene compounds is 5.8 Å as measured on Dreiding models, and the distance between these same carbons of the *cis* cyclohexane 14 is 4.8 Å. It should also be noted that rotation of the ring bearing the carboxylic acid about the 3-indole bond confers little change in the relative orientation of the carboxylic acid in the *trans* cyclohexane, cyclohexene, and benzene compounds, whereas the corresponding change is much greater in the *cis* cyclohexane compound 14. Of less importance, but contributing to enhanced potency, are the effects of N-alkylation of the indole and mono- or dicarboxylic acid functionality on the cyclohexanes. In six of seven cases tested, N-alkyl substituted compounds were significantly better inhibitors than the unsubstituted parents. Thus, 8, and 9 were more potent than 5. Similarly, 7 and 11 were more potent than 6, and 13 and 18 were more potent than 12 and 17, respectively. Only in the case of 10 was there a numerical but not statistically significant enhancement of activity over the unsubstituted 5. In the monocarboxylic acid series, N-Me substitution gave the most potent compound 8, while in the dicarboxylic acid series, activity of the N-Me compound 7 was not significantly different from the N-Et compound 11. The monocarboxylic acids 5, 8, 9, and 15 proved to be consistently more potent inhibitors than their

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dicarboxylic acid counterparts, 6, 7, 11, and 13, respectively. A parameter that did not seem to influence activity was the degree of saturation of the cyclohexane ring. Thus, 5 and 17 appeared to be equipotent, as were 15, 8, and 18.

Prior to developing the structural requirements of complement inhibitory activity in this series, a specific inhibitory site within the complement cascade was determined for compound 6 and this will be the subject of a separate publication. The relatively selective inhibitory activity of 6 (unpublished data) suggests that the "qualitative" SAR reported here may be the result of a specific reaction step. Alternately, it is possible that more than a single complement site has been altered, making the interpretation of these data more difficult.

The effects of these compounds (5–18) on the alternative complement pathway activity was also determined by hemolytic molecular titrations. None expressed significant activity (data not shown).

The reverse passive Arthus reaction (RPAR) in guinea pigs was chosen as a complement mediated model of acute inflammation to be used to identify compounds with in vivo complement inhibitory activity. Cobra venom factor (CVF) is known to suppress the RPAR by depleting complement levels in animals and it was used as reference material.¹³

The suppressive effects on the RPAR displayed by several of the in vitro complement inhibitors, when administered iv or ip, are reported in Table I. No test compound was active via the oral route (data not shown). Although in vivo suppression of the RPAR was established for this series of compounds, this activity was not consistent with the structural requirements that were determined in vitro. Not only were the in vitro active compounds 5, 6, 8, 15, 16, and 18 active in the RPAR but the inactive compound 14 also showed efficacy. Metabolism of 14 to a complement inhibitor is a possible explanation for this unexpected result. Alternatively, suppression of the RPAR might be via an unknown complement independent mechanism. Further investigations will be necessary to clarify this issue.

In summary, evidence has been presented supporting the contention that 4-(2-phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic acids possess unique structural features related to classical complement pathway inhibitory activity. These unique structural features include (a) orientation of the carboxylic acid in a trans stereochemistry rather than cis relative to the indole, (b) having the indole nitrogen substituted, and (c) placing one carboxylic acid function rather than two at the 1-position of the cyclohexane.

Experimental Section

Biology. C1–C9 Molecular Titrations. Hemolytic molecular titrations of classical complement activity were performed according to Rapp and Borsos¹¹ and complement inhibition assessed according to Hong et al.¹⁴ Operationally, drug was maintained in the presence of complement during the lytic reaction of sensitized erythrocytes. Equal volumes (0.2 mL) of normal human serum (NHS), diluted in GVB²⁺¹⁵ to effect lysis at $Z = 1$, and drug at various concentrations were mixed at 0 °C. The dilution of NHS effecting 63% lysis of optimally sensitized sheep erythrocyte (EA) was previously determined (1:100 to 1:200). EA (0.2 mL of 1.5×10^8 cells/mL) and 1.8 mL of GVB²⁺ were added to the

reaction mixture followed by incubating at 37 °C for 60 min. The extent of lysis was determined spectrophotometrically at 412 nm. The ratio of drug treated to untreated (Z/Z_0) reaction mixtures was determined as a function of drug concentration. The concentration of drug effecting 50% suppression ($Z/Z_0 = 0.5$ or IC_{50}) was determined.

Reverse Passive Arthus Reaction (RPAR). Male guinea pigs (250–300 g) were administered chicken egg albumin (CEA) intravenously (20 mg/kg) immediately before intradermal injection of 0.2 mL of goat anti-CEA containing 2.4 mg of Ab protein/mL. The hemorrhagic response was allowed to develop for 4 h, and the animals were sacrificed and skin lesions excised. The skin sections were homogenized, water extracted, and cleared, and the optical density at 541 or 412 nm was determined. A paired Student *t* test was used to estimate significant inhibition of drug-treated groups. *P* values less than 0.025 were considered significant. Drug was administered ip 2 h before eliciting the Arthus response and iv administration was effected 20 min before elicitation.

Chemistry. Microanalytical determinations were obtained on all new compounds reported and were carried out by Intranal Laboratories, Inc., Rensselaer, NY, and Galbraith Laboratories Inc., Knoxville, TN. Analyses for the indicated elements were within $\pm 0.4\%$ of the theoretical values. Melting points are uncorrected. ¹³C NMR spectra were determined on either a JEOL FX-60 60-MHz or JEOL FX-270 270-MHz instrument. Chemical shifts (δ) are reported in ppm relative to Me₄Si (δ 0.00, internal standard).

4-(2-Phenyl-1*H*-indol-3-yl)-3-cyclohexene-1-carboxylic Acid (5). A mixture of 100 g (0.52 mol) of 2-phenylindole, 80.8 g (0.57 mol) of 4-oxocyclohexanecarboxylic acid,¹⁶ 518 mL of AcOH, 103.6 mL of Ac₂O, and 25.9 mL of 85% H₃PO₄ was stirred at room temperature for 3 days and chilled in an ice bath to 20 °C. The precipitate that resulted was collected, washed with H₂O, pressed dry, and taken up in ether. This solution was washed with H₂O three times and with brine one time, dried (MgSO₄), charcoaled, evaporated to 200 mL, and let stand for 2 days to give a precipitate. The precipitate was collected and combined with a second crop to give 72.7 g (44%) of 5: mp 178–179 °C. A small portion was recrystallized from MeCN: mp 180–183 °C. Anal. (C₂₁H₁₉NO₂) C, H, N.

4-(2-Phenyl-1*H*-indol-3-yl)-3-cyclohexene-1,1-dicarboxylic Acid (6). By use of the method described for the preparation of 5, 105.9 g (0.57 mol) of 4-oxo-1,1-cyclohexanedicarboxylic acid¹⁶ was transformed into crude 6, which was slurried with 250 mL of boiling MeCN and collected to give 78 g (42%) of 6: mp 216 °C dec. A small portion was recrystallized from Et₂O: CHCl₂:hexane: mp 207–209 °C. Anal. (C₂₂H₁₉NO₄) C, H, N.

4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)-3-cyclohexene-1,1-dicarboxylic Acid (7). A mixture of 9.0 g (0.043 mol) of 1-methyl-2-phenylindole, 9.3 g (0.05 mol) of 4-oxo-1,1-cyclohexanedicarboxylic acid, 100 mL of AcOH, 4.7 mL of Ac₂O, and 1 mL of 85% H₃PO₄ was stirred at room temperature for 17 h, and then 100 mL of H₂O was added dropwise to give a white solid which was collected and washed with H₂O. The solid was taken up in Et₂O, washed with H₂O three times and with brine two times, dried (MgSO₄), concentrated in vacuo, and crystallized from MeCN to give 8.4 g (52%) of 7: mp 210–211 °C. Anal. (C₂₃H₂₁NO₄) C, H, N.

Methyl 4-(2-Phenyl-1*H*-indol-3-yl)-3-cyclohexene-1-carboxylate (5 Me Ester). A solution of 70.0 g (0.22 mol) of 5, 35.2 g (1.1 mol) of MeOH, 350 mL of CH₂Cl₂, and 0.5 mL of methanesulfonic acid was heated under reflux for 21 h. The reaction mixture was cooled, washed with H₂O, dried (Na₂SO₄), concentrated in vacuo, and crystallized from EtOAc:hexane to give 51.8 g (71%) of 5 Me ester in two crops: mp 175–177 °C. Anal. (C₂₂H₂₁NO₂) C, H, N.

Alternatively, 5 Me ester (mp 166–175 °C) was prepared in 81% yield from 31 g (0.098 mol) of 5, 47 g (0.34 mol) of powdered K₂CO₃, 47 mL (0.75 mol) of MeI, and 117 mL of DMF by using the method described for the preparation of dimethyl 4-(2-

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phenyl-1*H*-indol-3-yl)-3-cyclohexene-1,1-dicarboxylate (6 diMe ester).

Methyl 4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)-3-cyclohexene-1-carboxylate (8 Me Ester). A partial solution of 8.06 g (0.122 mol) of crushed KOH pellets in 100 mL Me₂SO was prepared by stirring the mixture for 45 min. 5 Me ester (12.0 g, 0.036 mol) was added in portions with stirring during 15 min to the partial solution. The resulting dark-red solution was stirred for 45 min, cooled in an ice bath, and treated dropwise with 10.2 g (0.072 mol) of MeI in 50 mL Me₂SO. Stirring was continued at room temperature for 3 h. The reaction mixture was cooled again in an ice bath and 250 mL of H₂O was added followed by Et₂O. The Et₂O layer was separated and the aqueous layer was extracted with Et₂O two times. The combined Et₂O solutions were washed with H₂O and brine, dried (MgSO₄), concentrated in vacuo, and crystallized from Et₂O:hexane to give 10.4 g of a solid (mp 122–125 °C). This solid was dissolved in CHCl₃ and the solution was charcoaled, concentrated in vacuo, and crystallized from CHCl₃:Et₂O:hexane two times to give 4.67 g (38%) of 8 Me ester: mp 135–136 °C. Anal. (C₂₃H₂₃NO₂) C, H, N.

4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)-3-cyclohexene-1-carboxylic Acid (8). To a solution of 5.6 g (0.085 mol) of KOH in 150 mL of EtOH was added 6.9 g (0.020 mol) of 8 Me ester. This mixture was heated under reflux until solution resulted. Heating was continued 4 h and then the solution was cooled, concentrated in vacuo, dissolved in H₂O, and made acidic with 2 N HCl to give a precipitate. The solid was collected, washed with H₂O, dissolved in Et₂O, washed with H₂O, dried (Na₂SO₄), charcoaled, concentrated in vacuo, and recrystallized twice from Et₂O:hexane to give 5.6 g (85%) of 8: mp 171–172 °C. Anal. (C₂₂H₂₁NO₂) C, H, N.

4-(1-Ethyl-2-phenyl-1*H*-indol-3-yl)-3-cyclohexene-1-carboxylic Acid (9). To 40 g (0.85 mol) of NaH (50% in oil) that was washed with pentane three times and stirred in 60 mL of DMF was added 12.2 g (0.38 mol) of 5 in portions. After the addition was complete, the mixture was heated at 50 °C for 2 h and then cooled in an ice bath, and 8 mL (0.10 mol) of EtI was added dropwise. The mixture was stirred at room temperature overnight and at 40 °C for 3 h. The reaction mixture was cooled, quenched with 250 mL of cold H₂O, and extracted with 500 mL of Et₂O. The Et₂O extract was washed with H₂O two times and brine one time, dried (MgSO₄), charcoaled, and concentrated in vacuo to give 13.5 g of an oil. This oil was combined with a solution of 2.6 g (0.04 mol) of KOH in 160 mL of EtOH and stirred at room temperature overnight. The reaction mixture was poured into 400 mL of cold H₂O, and the resulting mixture was washed with CH₂Cl₂ three times and Et₂O one time, filtered through filter cell, and made acidic with AcOH to give a precipitate. This precipitate was collected, washed with H₂O, and dried at 85 °C (0.1 mm) overnight to give 10.7 g (80%) of 9: mp 147–150 °C. Anal. (C₂₃H₂₃NO₂) C, H, N.

4-(2-Phenyl-1-propyl-1*H*-indol-3-yl)-3-cyclohexene-1-carboxylic Acid (10). By use of the method described for the preparation of 9, 15.0 g (0.047 mol) of 5 was transformed into 8.9 g (53%) of 10: mp 126–127 °C. Anal. (C₂₄H₂₅NO₂) C, H, N.

4-(1-Ethyl-2-phenyl-1*H*-indol-3-yl)-3-cyclohexene-1,1-dicarboxylic Acid (11). A mixture of 56 g (0.15 mol) of 6, 136 g (0.99 mol) of powdered K₂CO₃, and 369 mL of DMF was stirred at room temperature 2 h. MeI (146.8 mL, 2.08 mol) was added and the mixture was heated under reflux in a hot H₂O bath for 4 h. After stirring overnight at room temperature, the excess MeI was removed by concentrating the reaction mixture in vacuo at 30 °C. The mixture was then poured into 1200 mL of H₂O and extracted with EtOAc three times. The combined EtOAc extracts were washed with H₂O two times and brine one time, dried (MgSO₄), and concentrated in vacuo to give 63.1 g of the crude 6 diMe ester that was used in subsequent reactions without further purification.

To 1.8 g (0.37 mol) of NaH (50% in oil) that was washed three times with pentane and slurried and stirred in 50 mL of DMF was added 12 g (0.031 mol) of the crude 6 diMe ester in portions. After addition was complete, the mixture was heated at 50 °C for 2 h and then cooled in an ice bath, and 3.0 mL (0.048 mol) of EtI was added dropwise. The mixture was stirred at room temperature overnight and at 40 °C for 3 h. The reaction mixture was cooled, quenched with 300 mL of cold H₂O, and extracted

with Et₂O. The Et₂O extract was washed with H₂O two times and brine one time, dried (MgSO₄), and concentrated in vacuo to give 11.8 g of an oil which was combined with 4.3 g (0.065 mol) of KOH, 11.8 mL of H₂O, and 217 mL of EtOH, and heated under reflux for 5 h. The precipitate was collected (7.1 g), taken up in H₂O, filtered, and made acidic with 10% HCl to give a precipitate. This precipitate was collected, washed with H₂O, and dried at 70 °C (0.1 mm) overnight to give 5.7 g (49%) of 11: mp 160–165 °C. Anal. (C₂₄H₂₃NO₄) C, H, N.

4-(2-Phenyl-1*H*-indol-3-yl)cyclohexane-1,1-dicarboxylic Acid (12). A solution of 5.3 g (0.015 mol) of 6 in 100 mL of EtOH was combined with 1.2 g of 10% Pd/C and hydrogenated on a Parr apparatus at room temperature for 3 h. After the catalyst was removed, the solution was concentrated in vacuo to a gum. This gum was crystallized from Et₂O:hexane and then recrystallized three times from Et₂O:hexane to give 4.0 g (74%) of 12: mp 196–197 °C. Anal. (C₂₂H₂₁NO₄) C, H, N.

4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)cyclohexane-1,1-dicarboxylic Acid (13). By use of the method described for the preparation of 12, except that dioxane replaced EtOH as solvent, 7.5 g (0.020 mol) of 7 was hydrogenated to give 3.8 g (51%) of 13: mp 230 °C dec (after one crystallization from acetone:hexane). Anal. (C₂₃H₂₃NO₄) C, H, N.

cis-4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic Acid (14) and trans-4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)cyclohexane-1-carboxylic Acid (15). By use of the method described for the preparation of 13, 35 g (0.10 mol) of 8 Me ester was hydrogenated to give 35 g of a gum that was a mixture of two products (SiO₂ TLC, toluene eluate, *R*_f 0.3 and 0.4). The gum was triturated with Et₂O and the solid that formed was collected, washed with Et₂O, and dried to give 18.8 g of solid that was recrystallized from heptane to give 14.4 g (41%) of 14 Me ester with TLC *R*_f 0.4: mp 125–126 °C; ¹³C NMR (CDCl₃) δ 175.6 (COO), 137.2, 136.6, 132.4, 130.7, 128.1, 127.8, 126.2, 121.2, 120.3, 118.7, 118.1, 109.2, (indole and phenyl), 51.3 (CH₃O), 38.3 (CHCOO), 35.8 (indole CH), 30.4 (CH₃N), 29.1 (C₂ and C₆), 27.8 (C₃ and C₅). Anal. (C₂₃H₂₅NO₂) C, H, N.

The Et₂O filtrate was concentrated to a gum (16 g). This gum was dissolved in a minimum of toluene and subjected to HPLC chromatography on SiO₂ with toluene:hexane (9:1) as eluate to give a pure fraction as determined by TLC of the *R*_f 0.3 compound which was recrystallized from heptane to give 0.8 g of 15 Me ester: mp 141–142 °C; ¹³C NMR (CDCl₃) δ 176.4 (COO), 137.2, 137.0, 132.3, 130.6, 128.1, 127.9, 126.1, 121.2, 120.1, 118.6, 117.4, 109.4 (indole and phenyl), 51.3 (CH₃O), 42.9 (CHCOO), 35.7 (indole CH), 32.2 (C₂ and C₆), 30.4 (CH₃N), 29.4 (C₃ and C₅). Anal. (C₂₃H₂₅NO₂) C, H, N.

By use of the method described for the preparation of 8, 7 g (0.02 mol) of 14 Me ester was transformed into 5.1 g (77%) of 14: mp 270–271 °C (recrystallized from dioxane:H₂O); ¹³C NMR (Me₂SO-*d*₆) δ 175.7 (COO), 136.9, 136.2, 131.6, 130.2, 128.0, 127.8, 125.7, 120.7, 119.4, 118.1, 117.3, 109.6 (indole and phenyl), 37.5 (CHCOO), 35.3 (indole CH), 30.1 (CH₃N), 28.6 (C₂ and C₆), 27.3 (C₃ and C₅). Anal. (C₂₂H₂₃NO₂) C, H, N.

By use of the method described for the preparation of 8, 5.0 g (0.014 mol) of 15 Me ester was transformed into 2.8 g (60%) of 15: mp 178–180 °C (crystallized from Et₂O); ¹³C NMR (Me₂SO-*d*₆) δ 176.2 (COO), 136.8, 136.4, 131.7, 130.2, 128.0, 127.8, 125.7, 120.7, 119.6, 118.2, 116.7, 109.5 (indole and phenyl), 42.0 (CHCOO), 35.4 (indole CH), 31.6 (C₂ and C₆), 30.0 (CH₃N), 29.1 (C₃ and C₅). Anal. (C₂₂H₂₃N₂) C, H, N.

cis-4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)-1,1-cyclohexanedicarboxylic Acid Monomethyl Ester (16). To 1.8 g (0.037 mol) of NaH (50% in oil) that was washed with pentane three times and stirred in 50 mL of DMF was added dropwise 11.8 g (0.030 mol) of crude 6 diMe ester in 30 mL of DMF. After the addition was complete, the mixture was heated at 50 °C for 2 h and then cooled in an ice bath, and 3.0 mL (0.048 mol) of MeI was added dropwise. The mixture was stirred at room temperature overnight and at 50 °C for 3 h. The reaction mixture was poured into 300 mL of H₂O and extracted with Et₂O two times. The combined Et₂O extracts were washed with H₂O two times and brine one time, dried (MgSO₄), and concentrated in vacuo to give 10.6 g of resinous residue. By use of the method described for the preparation of 13, 10 g (0.025 mol) of the above resinous residue was hydrogenated to give 5.0 g (44% overall) of dimethyl

4-(1-methyl-2-phenyl-1*H*-indol-3-yl)-1,1-cyclohexanedicarboxylate (16 Me ester): mp 134–135 °C (crystallized from EtOH). Anal. (C₂₅H₂₇NO₄) C, H, N.

To a solution of 3.2 g (0.049 mol) of KOH in 50 mL of MeOH and 5 drops of H₂O was added 2.0 g (0.0049 mol) of 16 Me ester. This solution was heated under reflux for 1 h and then concentrated in vacuo at room temperature. The residue was suspended in H₂O, made acidic with 3 N HCl, and extracted with Et₂O. The Et₂O extract was washed with brine two times, dried (MgSO₄), concentrated in vacuo, and crystallized from Et₂O:hexane to give 0.8 g (42%) of 16: mp 192–193 °C. Anal. (C₂₄H₂₅NO₄) C, H, N.

4-(2-Phenyl-1*H*-indol-3-yl)benzoic Acid (17). To a solution of 43.4 g (0.13 mol) of 5 Me ester in 400 mL of dioxane was added 63.6 g (0.28 mol) of 2,3-dichloro-5,6-dicyanobenzoquinone (DDQ) followed by 50 mL of dioxane. The resulting pea-green suspension was stirred and heated under reflux for 19 h. The reaction mixture was cooled in an ice bath and the insoluble 2,3-dichloro-5,6-dicyanohydroquinone was collected, washed with hot dioxane:hexane, and dried to give 51.3 g (80%): mp 302–305 °C. The reaction mixture filtrate was concentrated and partitioned between EtOAc:toluene and ice H₂O. The organic layer was separated, dried (Na₂SO₄), and concentrated in vacuo to give 40 g of a semisolid. This semisolid was subjected to column chromatography on 1000 g of SiO₂ with toluene and toluene:EtOAc (9:1) as eluting solvents and gave 27.0 g of a red-orange solid: mp 163–165 °C. Purification of this material by HPLC on SiO₂ using hexane:toluene (1:4) as eluting solvent, followed by crystallization from EtOAc:hexane, gave 25.2 g (59%) of 17 Me ester: mp 165–166 °C. Anal. (C₂₂H₁₇NO₂) C, H, N.

A mixture of 13.2 g (0.040 mol) of 17 Me ester and 11.2 g (0.17 mol) of KOH in 250 mL of EtOH was stirred and heated under gentle reflux for 4 h to give a solution. The cooled solution was concentrated in vacuo and partitioned between cold H₂O and Et₂O, and the aqueous layer was separated, made acidic with 6 N HCl, and extracted with EtOAc. The EtOAc extract was dried (Na₂SO₄), concentrated in vacuo, and triturated with hexane to give a solid which was collected and recrystallized three times from acetone:EtOAc:hexane to give 8.9 g (71%) of 17: mp 252–253 °C. Anal. (C₂₁H₁₅NO₂) C, H, N.

4-(1-Methyl-2-phenyl-1*H*-indol-3-yl)benzoic Acid (18). By use of the method described for the preparation of 17, except that 18 was recrystallized from dioxane:H₂O, 10.2 g (0.030 mol) of 8 Me ester was transformed into 7.3 g (74%) of 18: mp 232–235 °C. Anal. (C₂₂H₁₇NO₂) C, H, N.

Acknowledgment. We thank Allan G. Hlavac for assistance in spectra determinations.

Registry No. 1, 948-65-2; 2, 3558-24-5; 3, 874-61-3; 4, 58230-12-9; 5, 93503-50-5; 5 Me ester, 93503-51-6; 6, 93503-52-7; 6 diMe ester, 93503-53-8; 7, 93503-54-9; 7 diMe ester, 93503-55-0; 8, 93503-56-1; 8 Me ester, 93503-57-2; 8-K, 93503-58-3; 9, 93503-59-4; 9 Me ester, 93503-60-7; 10, 93503-61-8; 11, 93503-62-9; 11 diMe ester, 93503-63-0; 12, 93503-64-1; 13, 93503-65-2; 14, 93503-66-3; 14 Me ester, 93503-67-4; 15, 93503-68-5; 15 Me ester, 93503-69-6; 16, 93503-70-9; 16 Me ester, 93503-71-0; 16 Me ester dihydro, 93503-55-0; 16-K, 93503-72-1; 17, 93503-73-2; 17 Me ester, 93503-74-3; 18, 93503-75-4; 2,3-dichloro-5,6-dicyanohydroquinone, 4640-41-9; EtI, 75-03-6; PrBr, 106-94-5; DDQ, 84-58-2.

Imidazo[1,5-*a*]pyridines: A New Class of Thromboxane A₂ Synthetase Inhibitors

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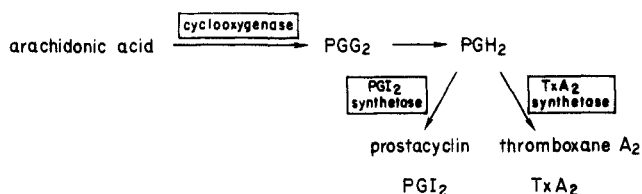
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Received December 7, 1983

The synthesis and structure-activity profile of a new class of potent and highly specific thromboxane A₂ synthetase inhibitors is described. The most potent member of this series in vitro is determined to be imidazo[1,5-*a*]pyridine-5-hexanoic acid (9).

Thromboxane A₂ (TxA₂) is an extremely unstable natural product with potent vasoconstricting, bronchoconstricting, and platelet aggregating activities.² It has been implicated in the etiology of a variety of disorders including vasospasm, stroke, ischemia, and myocardial infarction.³ The biosynthesis of TxA₂ from arachidonic acid is blocked by classical nonsteroidal antiinflammatory drugs which prevent the formation of its precursors prostaglandin G₂ and prostaglandin H₂ by inhibiting cyclooxygenase (Scheme I).

Since TxA₂ and prostacyclin (PGI₂) share PGG₂ as a common precursor, inhibition of cyclooxygenase necessarily blocks the formation of the antiaggregatory vasodilator, PGI₂. A selective inhibitor of TxA₂ synthetase, which has no inhibitory effect on prostacyclin synthetase, would appear to have therapeutic potential especially in the treatment of platelet-mediated disorders.⁴ Thromboxane synthetase inhibitors may have therapeutic utility in several conditions where platelets are believed to play a

Scheme I. Biosynthesis of Prostacyclin and Thromboxane A₂



role in the disease process, e.g., thromboembolic disorders,⁵ pulmonary hypertension,⁶ cardiac ischemia,⁷ endotoxin shock,⁸ and tumor metastasis.⁹

Imidazole and pyridine, and especially their N- and 3-substituted derivatives, respectively, are the only re-

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